

1 **Distribution of sex ratio distorters in natural populations of the isopod**

2 ***Armadillidium vulgare***

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16

17 **Abstract**

18

19 In the isopod *Armadillidium vulgare*, many females produce progenies with female-biased sex ratios,
20 due to two feminizing sex ratio distorters (SRD): *Wolbachia* endosymbionts and the *f* element. We
21 investigated the distribution and population dynamics of these SRD and mitochondrial DNA variation
22 in 16 populations from Europe and Japan. Confirming and extending results from the 1990's, we
23 found that the SRD are present at variable frequencies in populations, and that the *f* element is
24 overall more frequent than *Wolbachia*. The two SRD never co-occur at high frequency in any
25 population, suggesting an apparent mutual exclusion. We also detected *Wolbachia* or the *f* element
26 in some males, which likely reflects insufficient titer to induce feminization or presence of
27 masculinizing alleles. Our results are consistent with a single integration event of a *Wolbachia*
28 genome in the *A. vulgare* genome at the origin of the *f* element, which contradicts an earlier
29 hypothesis of frequent losses and gains. We identified strong linkage between *Wolbachia* strains and
30 mitochondrial haplotypes, but no association between the *f* element and mitochondrial background.
31 Our results open new perspectives on SRD evolutionary dynamics in *A. vulgare*, the evolution of
32 genetic conflicts and their impact on the variability of sex determination systems.

33

34 **Keywords**

35

36 Sex ratio distorter, endosymbiont, *Wolbachia*, *f* element, sex determination

37 1. Introduction

38

39 Sex ratio distorters (SRD) are selfish genetic elements located on sex chromosomes or transmitted by
40 a single sex, which skew the proportion of males and females in progenies towards the sex that
41 enhances their own vertical transmission [1]. Major SRD types include sex chromosome meiotic
42 drivers [2,3], B chromosomes [4], selfish mitochondria [5] and intracellular endosymbionts [6,7].
43 Collectively, they are found in a wide range of animal and plant species and they have had a
44 tremendous impact on the ecology and evolution of their host species [8,9]. One of the most
45 emblematic SRD is the bacterial endosymbiont *Wolbachia* [10,11]. *Wolbachia* is a cytoplasmic,
46 maternally inherited alpha-proteobacterium found in a wide range of arthropods and nematodes. In
47 arthropods, *Wolbachia* often manipulates host reproduction in favor of infected females, thereby
48 conferring itself a transmission advantage. This is achieved through various strategies, three of which
49 causing sex ratio distortions towards females: male killing, thelytokous parthenogenesis and
50 feminization of genetic males [6,7,10,11].

51 In the terrestrial isopod *Armadillidium vulgare*, chromosomal sex determination follows female
52 heterogamety (ZZ males and ZW females) [12–14]. However, many females produce progenies with
53 female-biased sex ratios, due to the presence of two feminizing SRD: *Wolbachia* endosymbionts and
54 a locus called the *f* element [6,15,16]. *Wolbachia* symbionts cause ZZ genetic males to develop as
55 phenotypic females [17]. Three *Wolbachia* strains have been described in *A. vulgare*, for which
56 feminization induction has been demonstrated (*wVulC* and *wVulM* strains [18,19]) or is strongly
57 suspected (*wVulP* strain [20]). The *f* element is a nuclear insert of a large portion of a feminizing
58 *Wolbachia* genome in the *A. vulgare* genome [21]. The *f* element induces female development, as a
59 W chromosome does, and it shows non-Mendelian inheritance, making it an SRD [21,22]. These SRD
60 may cause turnovers in sex determination mechanisms [6,15,23] and they could explain why sex
61 chromosome systems are so variable in terrestrial isopods [24–27].

62 Testing this hypothesis requires characterizing the evolutionary dynamics of SRD such as *Wolbachia*
63 and the *f* element in natural populations. In *A. vulgare*, this characterization is quite limited because
64 prior studies were mostly restricted to a narrow geographic area (western France), sometimes
65 focusing solely on *Wolbachia* [20,28–31]. The only exception is a 1993 study [32], which collated and
66 extended results from the early 1980's [33,34]. The main observations were that *Wolbachia* and the *f*
67 element are present at variable frequencies in field populations, and the *f* element is more frequent
68 than *Wolbachia*. However, earlier studies were limited by the lack of molecular tests for *Wolbachia*
69 and/or the *f* element, preventing any direct assessment of SRD presence. Instead, the authors used a

70 complex, indirect procedure combining a physiological test and crossings [32]. In addition to being
71 tedious and time-consuming (generation time is one year in this species), this procedure did not
72 allow direct and undisputable assessment of SRD presence. Moreover, it could only be run on
73 females and therefore provided no information on SRD presence in males. Finally, it could not reveal
74 individuals potentially carrying both SRD.

75 Here, we took advantage of the availability of molecular markers to directly assess SRD presence in
76 males and females from *A. vulgare* field populations from Europe and Japan. This approach allowed
77 us to circumvent the limitations of previous studies, and to revisit the population dynamics of
78 *Wolbachia* and the *f* element in this species and their association mitochondrial lineages.

79

80 **2. Materials and Methods**

81

82 647 *A. vulgare* individuals from 16 natural populations across Europe and Japan were collected by
83 hand. Individuals were sexed and stored in alcohol or at -20°C prior to DNA extraction. Total genomic
84 DNA was extracted from the head and legs of each individual, as described previously [21]. We used
85 four molecular markers to assess the presence of *Wolbachia* and the *f* element in DNA extracts: *Jtel*
86 [21], *wsp* [35], *recR* [36] and *ftsZ* [37]. While *Jtel* is specific to the *f* element, *wsp* and *recR* are specific
87 to *Wolbachia*, and *ftsZ* is present in both the *f* element and *Wolbachia* [21]. We assessed the
88 presence or absence of these markers by PCR, as described previously [21]. Different amplification
89 patterns were expected for individuals with *Wolbachia* only (*Jtel*⁻, *wsp*⁺, either *recR*⁺ or *ftsZ*⁺), the *f*
90 element only (*Jtel*⁺, *wsp*⁻, *recR*⁻), both *Wolbachia* and the *f* element present (*Jtel*⁺, *wsp*⁺, *recR*⁺) or
91 both *Wolbachia* and the *f* element lacking (*Jtel*⁻, *wsp*⁻, *ftsZ*⁻). The few individuals exhibiting other
92 amplification patterns were classified as “undetermined status”. A quantitative-PCR assay was used
93 to measure *Wolbachia* titer in some individuals (see supplementary Methods). To characterize
94 *Wolbachia* strain diversity, *wsp* PCR products were purified and Sanger sequenced using both
95 forward and reverse primers by GenoScreen (Lille, France). Forward and reverse reads were
96 assembled using Geneious® v.7.1.9 to obtain one consensus sequence per individual. To evaluate
97 mitochondrial diversity, we amplified by PCR a ~700 bp-long portion of the Cytochrome Oxidase I
98 (*COI*) gene in all individuals [38]. PCR products were purified and Sanger sequenced as described
99 above. Haplotype network analysis was performed using the *pegas* package [39]. All statistical
100 analyses were performed with R v.3.6.0 [40]. Figures were realized with *ggplot2* [41].

101

102 3. Results

103

104 We tested the presence of *Wolbachia* and the *f* element in 647 individuals (423 females and 224
105 males) from 16 populations across Europe and Japan (Tables 1, S1). While most males lacked both
106 SRD, 48% of females carried at least one of them. The remaining females presumably carry W
107 chromosomes, although the existence of other feminizing elements cannot be formally excluded. As
108 expected for feminizing elements, the SRD were mostly found in females, the *f* element being more
109 frequent than *Wolbachia* overall. Both SRD were found in the same individuals in only 3 females from
110 a single population (Chizé). *Wolbachia*-infected individuals carried one of the three previously known
111 *Wolbachia* strains of *A. vulgare*: wVulC (n=62), wVulM (n=23) or wVulP (n=4).

112 *Wolbachia* and *f* element distribution in females was highly heterogenous among populations (Figure
113 1a). These SRD were found in 10 and 11 out of 16 populations, but they reached frequencies >10% in
114 only 6 and 7 populations, respectively. The two SRD coexisted in 8 populations. A generalized linear
115 model predicting the frequency of the *f* element as a binomial response by the proportion of
116 individuals carrying *Wolbachia* (each statistical unit being a population) showed that the prevalence
117 of the two SRD was significantly negatively correlated (Chi-squared test, $p < 7.9 \times 10^{-8}$, 14 df) (Figure
118 1b). Hence, in Floirac, Poitiers, Saint Julien l'Ars and Pisa populations, *Wolbachia* was frequent (23-
119 94% frequency in females) and the *f* element was rare (0-8%). By contrast, the *f* element was
120 frequent (35-96%) and *Wolbachia* was rare (0-11%) in Prague, Beauvoir, Chizé, Coulombiers and La
121 Crèche populations. In the other populations, both SRD were found at low to moderate frequency (0-
122 19%), including 3 populations devoid of both SRD (Lastovo, Hyogo and Bucharest).

123 Males carrying *Wolbachia* or the *f* element were found in 2 and 4 out of 16 populations, respectively.
124 In all cases, these males occurred in populations in which the corresponding SRD were the most
125 prevalent ones in females: Beauvoir, Chizé, Coulombiers and La Crèche for the *f* element, and Floirac
126 and Saint Julien l'Ars for *Wolbachia*. Overall, these males had much lower *Wolbachia* titer than
127 females from their respective populations (Figure S1, Table S2).

128 The 642 individuals sequenced at the *COI* gene presented a total of 92 segregating sites defining 23
129 haplotypes (named I to XXIII; Table S3), with 1 to 7 haplotypes per population (Table S1, Figure 2).
130 The most frequent and widespread haplotype (I) was found in 188 individuals from 10 populations.
131 The second most frequent and widespread haplotype (V) was found in 106 individuals from 7
132 populations. We found 21 out of the 23 haplotypes among individuals lacking both *Wolbachia* and
133 the *f* element (Table 2, Figure 2). Among individuals carrying the *f* element, 6 haplotypes were found,
134 all but one (I, II, III, V and VI) being shared with individuals lacking both *Wolbachia* and the *f* element,

135 and one (IV) being carried by a single individual in the entire dataset. Among *Wolbachia*-infected
136 individuals, all those carrying *wVulC* were associated with either haplotype V or its close relatives (XI
137 and XII). All individuals carrying *wVulM* were associated with haplotype II and those carrying *wVulP*
138 with haplotype VII. Of the 5 haplotypes found in *Wolbachia*-infected individuals, 4 were shared with
139 individuals lacking both *Wolbachia* and the *f* element (II, V, VII and XII), 2 of which were also shared
140 with individuals carrying the *f* element (II and V), and one (XI) was present in a single individual in the
141 entire dataset.

142

143 **4. Discussion**

144

145 Our results provide direct evidence that the *f* element is overall more frequent than *Wolbachia* in the
146 sampled *A. vulgare* populations. We detected the *f* element in 11 *A. vulgare* natural populations from
147 4 European countries (Czech Republic, France, Germany and The Netherlands) and Japan. Together
148 with its previous detection in Denmark [21], our results indicate that the *f* element has spread to a
149 wide geographical range. The relative frequencies of the *f* element and *Wolbachia* are highly variable
150 among populations and, in general, when one SRD is frequent, the other SRD is rare. Overall, these
151 results are consistent with earlier results from the 1990's [32], although no molecular assay allowing
152 direct testing was available at that time and SRD presence or absence was inferred indirectly.

153 As the *JteI* marker is located across the site of integration of the *f* element in the *A. vulgare*
154 chromosome [21], we may conclude that *f* element presence in various populations results from a
155 single event of integration of a *Wolbachia* genome in the *A. vulgare* genome. An alternative scenario
156 would require independent insertions at the same chromosomal site, which is highly unlikely. This
157 conclusion contradicts an earlier hypothesis on the evolutionary dynamics of the *f* element, which
158 suggested that the *f* element was unstably integrated in the *A. vulgare* genome, experiencing
159 frequent loss from oocytes and recurrent gain from *Wolbachia* endosymbionts [22,23,42–44]. Under
160 this scenario, multiple independent *f*-like elements would be expected to segregate at low
161 frequencies in populations and they should be integrated in different genomic locations [16]. While
162 our results do not formally invalidate the possibility of additional *f*-like integrations in *A. vulgare*
163 populations, which the *JteI* marker would not detect, all observations can parsimoniously be
164 explained by a single origin of the *f* element. Examination of sex ratios from progenies of wild-caught
165 females lacking both SRD may offer further insight into this issue.

166 Using molecular assays allowed us to circumvent two limitations of the previously used physiological
167 test: the impossibility to detect *Wolbachia* and the *f* element in males, and the impossibility to detect
168 individuals carrying both SRD. Regarding *Wolbachia* presence in males, the historic protocol was only
169 applicable to females per design [29,30,32] and subsequent PCR screens for *Wolbachia* infection
170 have mostly focused on testing females [20,30,31]. In fact, males have seldom been tested and found
171 to carry *Wolbachia* [45]. Here, we detected *Wolbachia* in 7 males from 2 populations (Florac and
172 Saint Julien l’Ars), carrying either *wVulC* or *wVulM* strains. The failure of feminization by *Wolbachia*
173 most certainly reflects insufficient bacterial titer to induce feminization (Figure S1). These field
174 observations hence support the view that titer is an important factor for successful feminization, as
175 low titer is linked to incomplete feminization and intersexual phenotypes [42,46].

176 We also detected the presence of the *f* element in 11 males from 4 populations. Historically, the
177 presence of the *f* element in males has been indirectly inferred from crossings and the resulting sex-
178 ratios biases of progenies [22,43,47]. Our results constitute the first direct evidence for the presence
179 of the *f* element in *A. vulgare* males. In all 4 populations in which *f*-carrying males were found, the *f*
180 element was also frequent in females. Altogether, these observations suggest that the 11 males
181 carrying the *f* element also carry the masculinizing dominant allele known as “*M*” [16,43,47]. Indeed,
182 the *M* allele is able to restore a male phenotype in individuals carrying the *f* element [16,43,47].
183 Moreover, the *M* allele is thought to have been selected in response to female-biased sex ratios
184 caused by the *f* element [47]. Thus, the *M* allele is expected to rise in frequency when the *f* element
185 is frequent in a population [47], which is consistent with our observations. Unfortunately, no
186 molecular marker of the *M* allele is currently available, which prevents any direct assessment of its
187 actual presence in these populations. Thus, we cannot exclude that males carrying the *f* element
188 simply carry non-feminizing variants of this SRD.

189 Our results show that *Wolbachia* and the *f* element never co-occur at high frequency in any
190 population. This apparent mutual exclusion can be explained considering that co-occurrence of
191 multiple feminizing factors in a population should favor the most transmitted one [16,48]. Hence,
192 *Wolbachia* is expected to lead to the loss of nuclear feminizing elements in *A. vulgare* populations.
193 This situation does not result from an interference between chromosomes and *Wolbachia* within
194 individuals, but from counter selection of nuclear feminizing alleles in a population that becomes
195 increasingly biased towards females. Hence, the rise of *Wolbachia* would associate with the decline
196 of the *f* element in a population. Why, under these circumstances, *Wolbachia* has not invaded all *A.*
197 *vulgare* populations is still unclear and may reflect fitness effects or possible resistance genes.

198 As a result, only very few individuals were found to carry both *Wolbachia* and the *f* element. They
199 represent only 3 females, all from the Chizé population (Figure 1a). These were likely born from
200 mothers carrying *Wolbachia* and fathers carrying the *f* element, which are frequent at Chizé. The
201 apparent absence of carriers of both SRD in other populations where these SRD are present could
202 simply be explained by the paucity of males carrying the *f* element.

203 Mapping SRD distribution onto mitochondrial genealogy showed excellent congruence between
204 *Wolbachia* strains and mitochondrial haplotypes (*wVulC-V*, *wVulM-II* and *wVulP-VII*). Such strong
205 association has previously been noted in *A. vulgare-Wolbachia* interactions at a smaller geographic
206 scale [30,31] and, more generally, in many arthropod-*Wolbachia* interactions [49]. This result
207 corroborates the rarity of non-maternal transmission of *Wolbachia* in *A. vulgare*. By contrast, the *f*
208 element was found in 6 different mitochondrial backgrounds (I-VI) scattered across the
209 mitochondrial phylogeny, indicating no particular association between the *f* element and
210 mitochondria. This result confirms and extends earlier data focused on western France and in which *f*
211 element presence in females was indirectly inferred based on sex ratios of their progenies [30]. This
212 observation can be explained by the occasional paternal transmission of the *f* element, which breaks
213 its association with mitochondrial background [16,22,30].

214

215 **Data accessibility**

216

217 All data are provided in the electronic supplementary material.

218

219

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221

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227

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231

232 **Figure legends**

233

234 **Figure 1. (A)** Prevalence of *Wolbachia* and the *f* element in males (m) and females (f) from 16
235 *Armadillidium vulgare* populations. **(B)** Relative proportions of *Wolbachia* and the *f* element in 16 *A.*
236 *vulgare* populations (represented by open circles).

237

238 **Figure 2.** Haplotype network of 23 mitochondrial variants (I-XXIII) from 16 *Armadillidium vulgare*
239 populations. Each circle represents one haplotype and circle size is proportional to the number of
240 individuals carrying the haplotype. Branch lengths connecting circles are proportional to divergence
241 between haplotypes. Sex ratio distorter frequencies are color-coded for each haplotype.

242

243 **Table 1. Prevalence of *Wolbachia* and *f* element sex ratio distorters in 16 populations of *Armadillidium vulgare*.**

244

Population	Country	Sample size	Sex	Number of individuals	No <i>f</i> element, no <i>Wolbachia</i>	Only <i>f</i> element	Only <i>Wolbachia</i>				Both wVulM and <i>f</i> element	Undetermined status
							wVulC	wVulM	wVulP	Undetermined		
Lastovo	Croatia	54	Males	30	30							
			Females	24	24							
Prague	Czech Republic	36	Males	9	9							
			Females	27	1	26						
Beauvoir	France	31	Males	6	5	1						
			Females	25	9	14		1			1	
Chizé	France	52	Males	8	2	6						
			Females	44	3	36		2		3		
Coulombiers	France	24	Males	4	2	2						
			Females	20	6	13	1					
Floirac	France	114	Males	38	34		2				2	
			Females	76	21	6	40	9				
Gript	France	45	Males	15	15							
			Females	30	26	2	2					
La Crèche	France	58	Males	21	19	2						
			Females	37	23	13	1					
Poitiers	France	23	Males	4	4							
			Females	19	10	1		4	4			
Saint Julien l'Ars	France	31	Males	14	9		1	3		1		
			Females	17	1		12	3				
Göttingen	Germany	24	Males	7	3						1	
			Females	17	11	3		2			4	
Pisa	Italy	28	Males	15	15							
			Females	13	10		3					
Hyogo	Japan	50	Males	21	18						3	

Tottori	Japan	49	Females	29	26						3	
			Males	21	21							
Bucharest	Romania	17	Females	28	26	2						
			Males	9	9							
Wageningen	The Netherlands	11	Females	8	8							
			Males	2	2							
			Females	9	7	1						1
Total males				224	197	11	3	3	1		9	
Total females				423	212	117	59	17	4	4	3	7
Total				647	409	128	62	20	4	5	3	16

245

246

247 **Table 2. Distribution of mitochondrial haplotypes in 642 *Armadillidium vulgare* individuals from 16 populations.**

248

Sex ratio distorter status	Number of individuals	Haplotype number	Haplotype list
No <i>f</i> element, no <i>Wolbachia</i>	404	21	I, II, III, V, VI, VII, VIII, IX, X, XII, XIII, XIV, XV, XVI, XVII, XVIII, XIX, XX, XXI, XXII, XXIII
<i>f</i> element only	128	6	I, II, III, IV, V, VI
<i>Wolbachia</i> (<i>wVulC</i> strain) only	62	3	V, XI, XII
<i>Wolbachia</i> (<i>wVulM</i> strain) only	20	1	II
<i>Wolbachia</i> (<i>wVulP</i> strain) only	4	1	VII
<i>Wolbachia</i> (undetermined strain) only	5	2	II, VII
Both <i>wVulM</i> and <i>f</i> element	3	1	II
Undetermined status	16	4	I, V, VI, XIX

249

250

251 **References**

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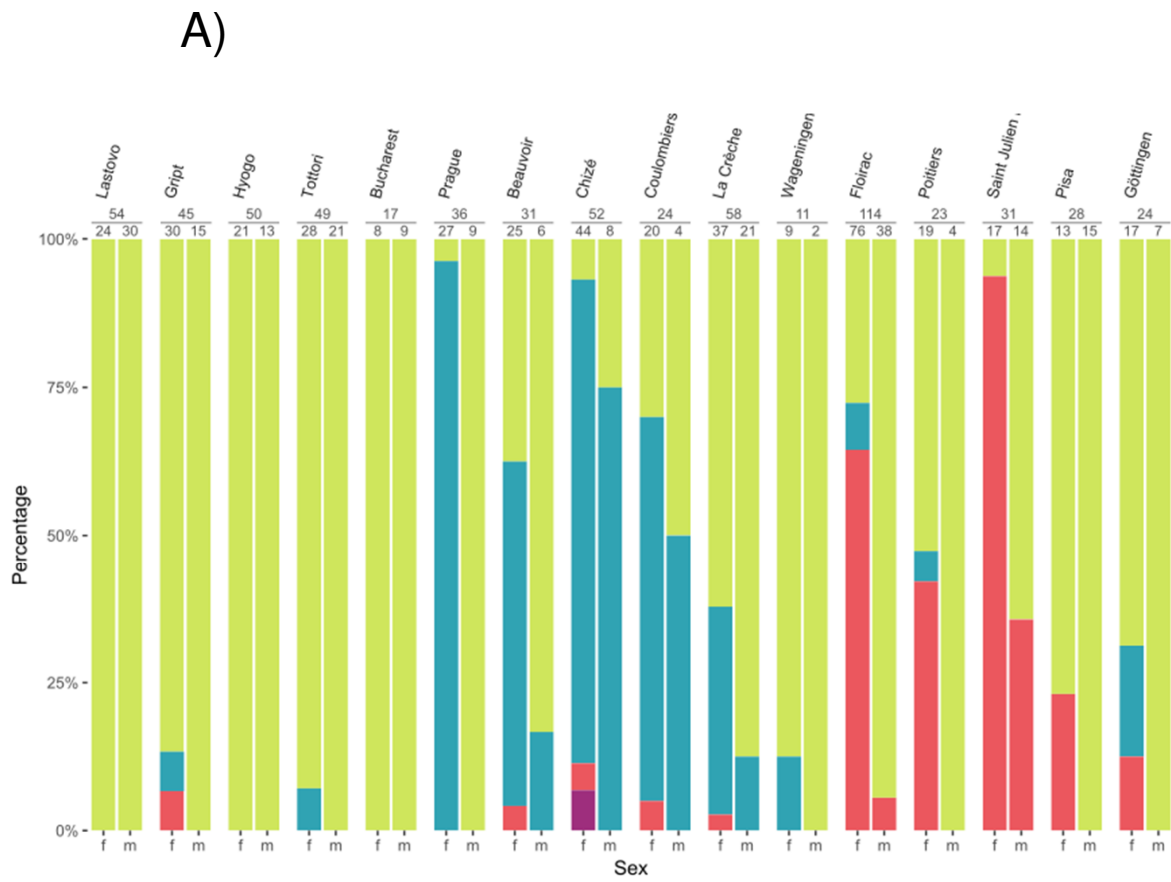
- 253 1. Beukeboom LW, Perrin N. 2014 *The evolution of sex determination*. Oxford: Oxford University
254 Press.
- 255 2. Jaenike J. 2001 Sex chromosome meiotic drive. *Annu. Rev. Ecol. Syst.* **32**, 25–49.
256 (doi:10.1146/annurev.ecolsys.32.081501.113958)
- 257 3. Helleu Q, Gérard PR, Montchamp-Moreau C. 2014 Sex chromosome drive. *Cold Spring Harb.*
258 *Perspect. Biol.* **7**, a017616. (doi:10.1101/cshperspect.a017616)
- 259 4. Camacho JPM, Schmid M, Cabrero J. 2011 B chromosomes and sex in animals. *Sex. Dev. Genet.*
260 *Mol. Biol. Evol. Endocrinol. Embryol. Pathol. Sex Determ. Differ.* **5**, 155–166.
261 (doi:10.1159/000324930)
- 262 5. Chase CD. 2007 Cytoplasmic male sterility: a window to the world of plant mitochondrial-nuclear
263 interactions. *Trends Genet.* **23**, 81–90. (doi:10.1016/j.tig.2006.12.004)
- 264 6. Cordaux R, Bouchon D, Greve P. 2011 The impact of endosymbionts on the evolution of host sex-
265 determination mechanisms. *Trends Genet* **27**, 332–41.
- 266 7. Hurst GDD, Frost CL. 2015 Reproductive parasitism: maternally inherited symbionts in a biparental
267 world. *Cold Spring Harb. Perspect. Biol.* **7**, a017699. (doi:10.1101/cshperspect.a017699)
- 268 8. Burt A, Trivers R. 2006 *Genes in conflict*. Cambridge, Massachusetts: The Belknap Press of Harvard
269 University Press.
- 270 9. Werren JH. 2011 Selfish genetic elements, genetic conflict, and evolutionary innovation. *Proc.*
271 *Natl. Acad. Sci. U. S. A.* **108 Suppl 2**, 10863–10870. (doi:10.1073/pnas.1102343108)
- 272 10. Werren JH, Baldo L, Clark ME. 2008 Wolbachia: master manipulators of invertebrate biology.
273 *Nat Rev Microbiol* **6**, 741–51. (doi:10.1038/nrmicro1969)
- 274 11. Kaur R, Shropshire JD, Cross KL, Leigh B, Mansueto AJ, Stewart V, Bordenstein SR,
275 Bordenstein SR. 2021 Living in the endosymbiotic world of Wolbachia: A centennial review. *Cell*
276 *Host Microbe* **29**, 879–893. (doi:10.1016/j.chom.2021.03.006)
- 277 12. Juchault P, Legrand JJ. 1972 Croisement de néo-mâles expérimentaux chez *Armadillidium*
278 *vulgare* Latr. (Crustace, Isopode, Oniscoïde). Mise en évidence d'une hétérogamétie femelle. *C R*
279 *Acad Sci Paris* **274**, 1387–1389.
- 280 13. Chebbi MA, Becking T, Moumen B, Giraud I, Gilbert C, Peccoud J, Cordaux R. 2019 The
281 Genome of *Armadillidium vulgare* (Crustacea, Isopoda) Provides Insights into Sex Chromosome
282 Evolution in the Context of Cytoplasmic Sex Determination. *Mol. Biol. Evol.* **36**, 727–741.
283 (doi:10.1093/molbev/msz010)
- 284 14. Cordaux R, Chebbi MA, Giraud I, Pleydell DRJ, Peccoud J. 2021 Characterization of a Sex-
285 Determining Region and Its Genomic Context via Statistical Estimates of Haplotype Frequencies in
286 Daughters and Sons Sequenced in Pools. *Genome Biol. Evol.* **13**, evab121.
287 (doi:10.1093/gbe/evab121)

- 288 15. Rigaud T, Juchault P, Mocquard JP. 1997 The evolution of sex determination in isopods
289 crustaceans. *Bioessays* **19**, 409–416.
- 290 16. Cordaux R, Gilbert C. 2017 Evolutionary Significance of Wolbachia-to-Animal Horizontal Gene
291 Transfer: Female Sex Determination and the f Element in the Isopod *Armadillidium vulgare*. *Genes*
292 **8**, 186. (doi:10.3390/genes8070186)
- 293 17. Martin G, Juchault P, Legrand JJ. 1973 Mise en évidence d'un micro-organisme
294 intracytoplasmique symbiote de l'Oniscoïde *Armadillidium vulgare* L. dont la présence
295 accompagne l'intersexualité ou la féminisation totale des mâles génétiques de la lignée thélygène.
296 *Comptes Rendus Académie Sci. Paris* **276**, 2313–2316.
- 297 18. Rigaud T, Souty Grosset C, Raimond R, Mocquard JP, Juchault P. 1991 Feminizing
298 endocytobiosis in the terrestrial crustacean *Armadillidium vulgare* Latr. (Isopoda): Recent
299 acquisitions. *Endocytobiosis Cell Res* **7**, 259–273.
- 300 19. Cordaux R, Michel-Salzat A, Frelon-Raimond M, Rigaud T, Bouchon D. 2004 Evidence for a
301 new feminizing Wolbachia strain in the isopod *Armadillidium vulgare*: evolutionary implications.
302 *Heredity* **93**, 78–84. (doi:10.1038/sj.hdy.6800482)
- 303 20. Verne S, Johnson M, Bouchon D, Grandjean F. 2007 Evidence for recombination between
304 feminizing Wolbachia in the isopod genus *Armadillidium*. *Gene* **397**, 58–66.
305 (doi:10.1016/j.j.gene.2007.04.006)
- 306 21. Leclercq Sb, Thézé J, Chebbi MA, Giraud I, Moumen B, Ernenwein L, Greve P, Gilbert C,
307 Cordaux R. 2016 Birth of a W sex chromosome by horizontal transfer of *Wolbachia* bacterial
308 symbiont genome. *Proc. Natl. Acad. Sci. U. S. A.* **113**, 15036–15041.
309 (doi:10.1073/pnas.1608979113)
- 310 22. Legrand JJ, Juchault P. 1984 Nouvelles données sur le déterminisme génétique et
311 épigénétique de la monogénie chez le crustacés isopodes terrestres *Armadillidium vulgare* Latr.
312 *Génét Sél Evol* **16**, 57–84.
- 313 23. Juchault P, Mocquard JP. 1993 Transfer of a parasitic sex factor to the nuclear genome of the
314 host: A hypothesis on the evolution of sex-determining mechanisms in the terrestrial isopod
315 *Armadillidium vulgare* Latr. *J Evol Biol* **6**, 511–528.
- 316 24. Juchault P, Rigaud T. 1995 Evidence for female heterogamety in two terrestrial crustaceans
317 and the problem of sex chromosome evolution in isopods. *Heredity* **75**, 466–471.
- 318 25. Becking T, Giraud I, Raimond M, Moumen B, Chandler C, Cordaux R, Gilbert C. 2017 Diversity
319 and evolution of sex determination systems in terrestrial isopods. *Sci. Rep.* **7**, 1–14.
320 (doi:10.1038/s41598-017-01195-4)
- 321 26. Becking T, Chebbi MA, Giraud I, Moumen B, Laverré T, Caubet Y, Peccoud J, Gilbert C,
322 Cordaux R. 2019 Sex chromosomes control vertical transmission of feminizing Wolbachia
323 symbionts in an isopod. *PLOS Biol.* **17**, e3000438. (doi:10.1371/journal.pbio.3000438)
- 324 27. Russell A, Borrelli S, Fontana R, Laricchiuta J, Pascar J, Becking T, Giraud I, Cordaux R,
325 Chandler CH. 2021 Evolutionary transition to XY sex chromosomes associated with Y-linked
326 duplication of a male hormone gene in a terrestrial isopod. *Heredity* **127**, 266–277.
327 (doi:10.1038/s41437-021-00457-2)

- 328 28. Juchault P, Legrand JJ, Mocquard JP. 1980 Contribution à l'étude qualitative et quantitative
329 des facteurs contrôlant le sexe dans les populations du crustacé isopode terrestre *Armadillidium*
330 *vulgare* Latreille. I. La population de Niort (Deux Sèvres). *Arch Zool Exp Gen* **121**, 3–27.
- 331 29. Grandjean F, Rigaud T, Raimond R, Juchault P, Souty-Grosset C. 1993 Mitochondrial DNA
332 polymorphism and feminizing sex factor dynamics in a natural population of *Armadillidium*
333 *vulgare* (Crustacea, Isopoda). *Genetica* **92**, 55–60.
- 334 30. Rigaud T, Bouchon D, Souty-Grosset C, Raimond R. 1999 Mitochondrial DNA polymorphism,
335 sex ratio distorters and population genetics in the isopod *Armadillidium vulgare*. *Genetics* **152**,
336 1669–1677.
- 337 31. Verne S, Johnson M, Bouchon D, Grandjean F. 2012 Effects of parasitic sex-ratio distorters on
338 host genetic structure in the *Armadillidium vulgare*-*Wolbachia* association. *J. Evol. Biol.* **25**, 264–
339 76. (doi:10.1111/j.1420-9101.2011.02413.x)
- 340 32. Juchault P, Rigaud T, Mocquard JP. 1993 EVOLUTION OF SEX DETERMINATION AND SEX-
341 RATIO VARIABILITY IN WILD POPULATIONS OF ARMADILLIDIUM-VULGARE (LATR) (CRUSTACEA,
342 ISOPODA) - A CASE-STUDY IN CONFLICT-RESOLUTION. *Acta Oecologica-Int. J. Ecol.* **14**, 547–562.
- 343 33. Juchault P, Legrand JJ. 1981 Contribution à l'étude qualitative et quantitative des facteurs
344 contrôlant le sexe dans les populations du Crustacé Isopode terrestre *Armadillidium vulgare* Latr.
345 II - Populations hébergeant le facteur féminisant F (bactérie intracytoplasmique). *Arch Zool Exp*
346 *Gén* **122**, 65–74.
- 347 34. Juchault P, Legrand JJ. 1981 Contribution à l'étude qualitative et quantitative des facteurs
348 contrôlant le sexe dans les populations du Crustacé isopode terrestre *Armadillidium vulgare*
349 Latreille. III. Populations n'hébergeant pas le facteur féminisant F (bactéroïde intracytoplasmique).
350 *Arch Zool Exp Gén* **122**, 117–131.
- 351 35. Braig HR, Zhou WG, Dobson SL, O'Neill SL. 1998 Cloning and characterization of a gene
352 encoding the major surface protein of the bacterial endosymbiont *Wolbachia pipientis*. *J.*
353 *Bacteriol.* **180**, 2373–2378.
- 354 36. Badawi M, Giraud I, Vavre F, Grève P, Cordaux R. 2014 Signs of Neutralization in a Redundant
355 Gene Involved in Homologous Recombination in *Wolbachia* Endosymbionts. *Genome Biol. Evol.* **6**,
356 2654–2664. (doi:10.1093/gbe/evu207)
- 357 37. Werren JH, Zhang W, Guo LR. 1995 Evolution and phylogeny of *Wolbachia*: reproductive
358 parasites of arthropods. *Proc. Biol. Sci.* **261**, 55–63. (doi:10.1098/rspb.1995.0117)
- 359 38. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994 DNA primers for amplification of
360 mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar.*
361 *Biol. Biotechnol.* **3**, 294–299.
- 362 39. Paradis E. 2010 pegas: an R package for population genetics with an integrated–modular
363 approach. *Bioinformatics* **26**, 419–420.
- 364 40. R Development Core Team. 2013 *R: A language and environment for statistical computing*.
365 See <http://www.R-project.org/>.
- 366 41. Wickham H *et al.* 2020 ggplot2: Create Elegant Data Visualisations Using the Grammar of
367 Graphics.

- 368 42. Juchault P, Legrand JJ. 1989 Sex determination and monogeny in terrestrial isopods
369 *Armadillidium vulgare* (Latreille, 1804) and *Armadillidium nasatum* bundde-lund, 1885. *Monit.*
370 *Zool Ital NS Monogr* **4**, 359–375.
- 371 43. Juchault P, Rigaud T, Mocquard J-P. 1992 Evolution of sex-determining mechanisms in a wild
372 population of *Armadillidium vulgare* Latr. (Crustacea, Isopod) : competition between two
373 feminizing parasitic sex factors. *Heredity* **69**, 382–390.
- 374 44. Rigaud T, Mocquard J-P, Juchault P. 1992 The spread of parasitic sex factors in populations of
375 *Armadillidium vulgare* Latr. (Crustacea, Oniscidae): effects on sex ratio. *Génét Sél Evol* **24**, 3–18.
- 376 45. Dittmer J, Lesobre J, Moumen B, Bouchon D. 2016 Host origin and tissue microhabitat
377 shaping the microbiota of the terrestrial isopod *Armadillidium vulgare*. *FEMS Microbiol. Ecol.* **92**,
378 fiw063. (doi:10.1093/femsec/fiw063)
- 379 46. Legrand JJ, Juchault P. 1986 Rôle des bactéries symbiotiques dans l'intersexualité, la
380 monogénie et la spéciation chez les crustacés oniscoïdes. *Boll Zool* **53**, 161–172.
- 381 47. Rigaud T, Juchault P. 1993 Conflict between feminizing sex ratio distorters and an autosomal
382 masculinizing gene in the terrestrial isopod *Armadillidium vulgare* Latr. *Genetics* **133**, 247–252.
- 383 48. Taylor DR. 1990 Evolutionary consequences of cytoplasmic sex ratio distorters. *Evol. Ecol.* **4**,
384 235–248.
- 385 49. Galtier N, Nabholz B, Glémin S, Hurst GDD. 2009 Mitochondrial DNA as a marker of molecular
386 diversity: a reappraisal. *Mol. Ecol.* **18**, 4541–4550. (doi:10.1111/j.1365-294X.2009.04380.x)
- 387

Figure 1



B)

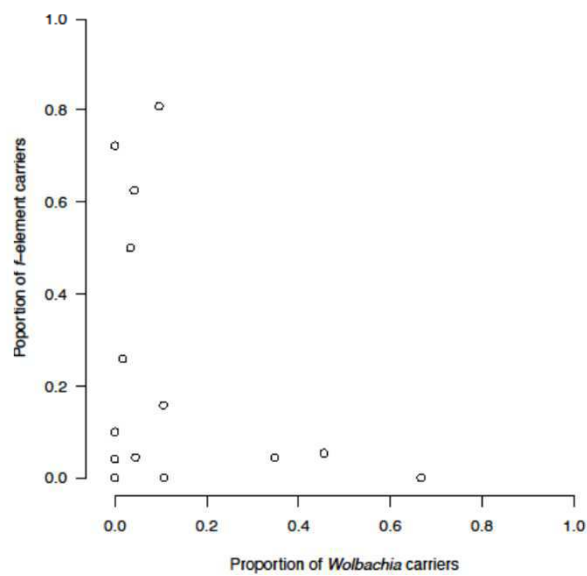


Figure 2

